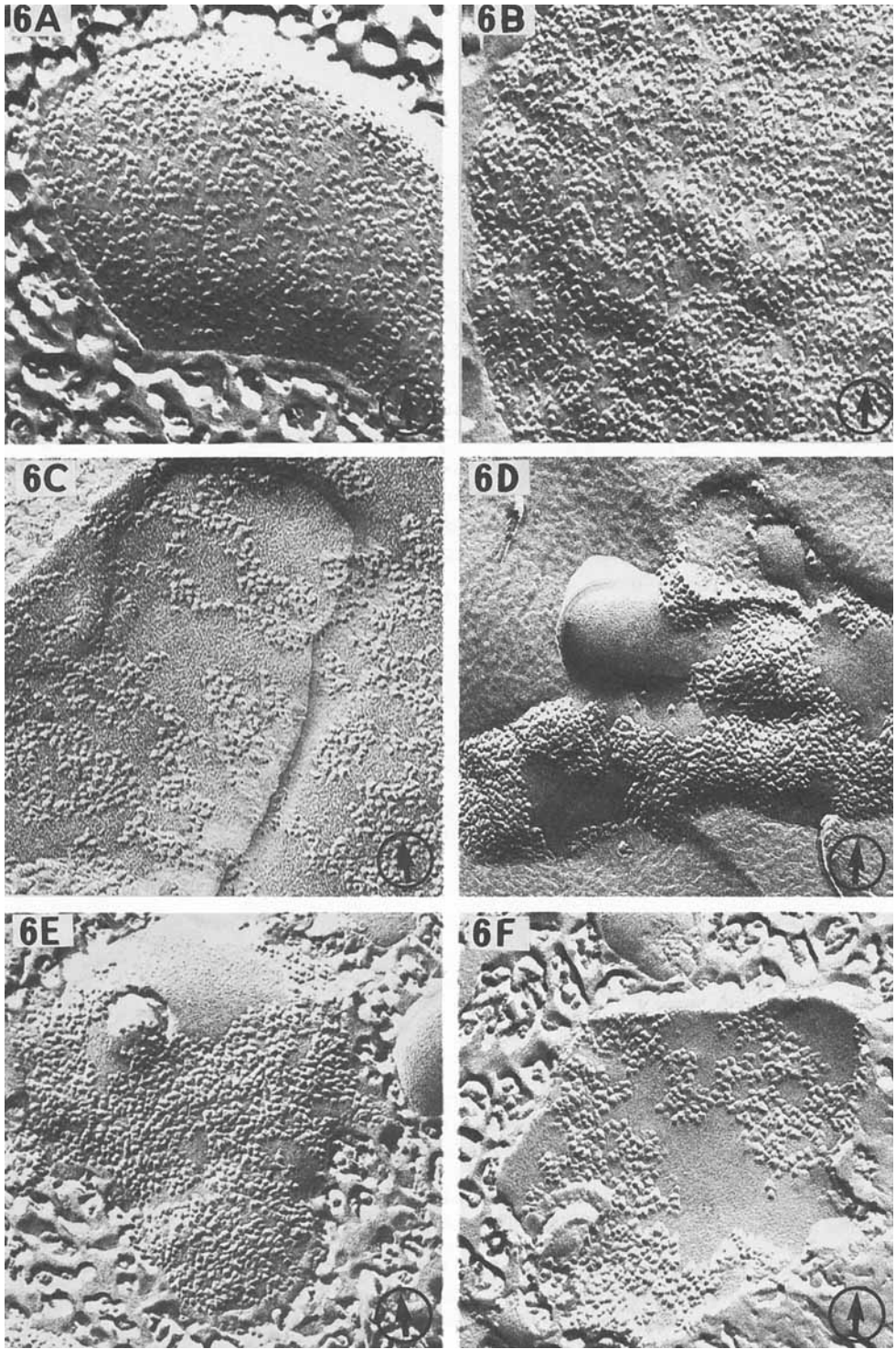
PRINTER'S CORRECTION

Because of unsatisfactory reproduction of three figures in **Journal of Supramolecular Structure, Volume 8, Number 3, 1978,** they are being reprinted here.

Figure 6 is from **Distribution of Glycophorin on the Surface of Human Erythrocyte Membranes and Its Association With Intramembrane Particles: An Immunochemical and Freeze-Fracture Study of Normal and En(a—) Erythrocytes,** Carl G. Gahmberg, Georg Taurén, Ismo Virtanen, and Jorma Wartiovaara, pages 337–347. This figure is reprinted on page 144, and the legend for it, at the bottom of this page.

Figures 2 and 7 are from **Vesicles Isolated From ATP-Depleted Erythrocytes and out of Thrombocyte-Rich Plasma,** Hans U. Lutz, pages 375–389. Figure 2 is reprinted on page 145, figure 7 on page 146.

Fig 6. Cleaved fracture faces of normal and En(a–) erythrocytes. Both in membranes from normal (A) and En(a–) (B) erythrocytes a similar even distribution of the intramembrane particles is seen. When membranes of normal cells were depleted of spectrin by incubation in 20 mM sodium phosphate (pH 8.5) at 37°C for 16h and then transferred to 0.05 M sodium phosphate (pH 6.0) and incubated at 37°C for 30 min, a slight aggregation of IMPs is seen (C). After a similar treatment of En(a–) membranes a clear aggregation is seen (D). When membranes were depleted of spectrin by incubation in 0.1 mM EDTA (pH 8.0) at 4°C for 16 h and then incubated in PBS at 37°C for 30 min, an extensive aggregation of the IMPs occurred both in normal (E) and En(a–) (F) membranes. Shadowing direction is indicated by circled arrow.(Magnification 75,000 ×.)


6A
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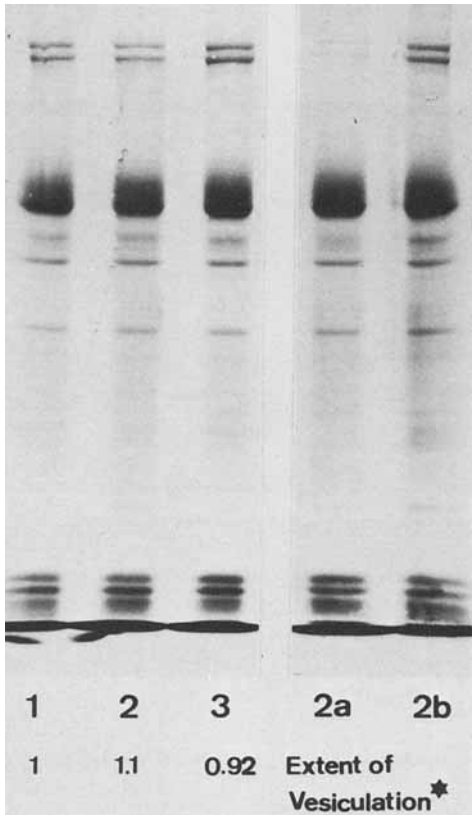

Fig 2. Extent of vesiculation and protein composition of vesicles released from ATP-depleted human erythrocytes in the presence of Ca^{+2} or EGTA. Erythrocytes were washed as outlined in Methods and surface-labeled with ^{125}I [12] prior to incubation. ATP-depletions were carried out for 38 hours in the medium given in Methods supplemented with 5 mM phosphate and with no further addition in experiment 1, with 50 μM $CaCl_2$ in experiment 2, and with 50 μM EGTA in experiment 3. 2a refers to the light vesicle fraction; 2b, to the heavy vesicle fractions.

*Extents of vesiculation are listed as relative numbers based on the number of cpm from ^{125}I recovered in the vesicle pellet originating from 10 ml of supernate from ATP-depleted cells. Extent 1, listed for the control (experiment 1), corresponds to 16,712 cpm recovered in the vesicle pellet. Extent data shown for experiments 1 and 2 are averages from two separate experiments. Radioactivity and protein were determined in triplicate.

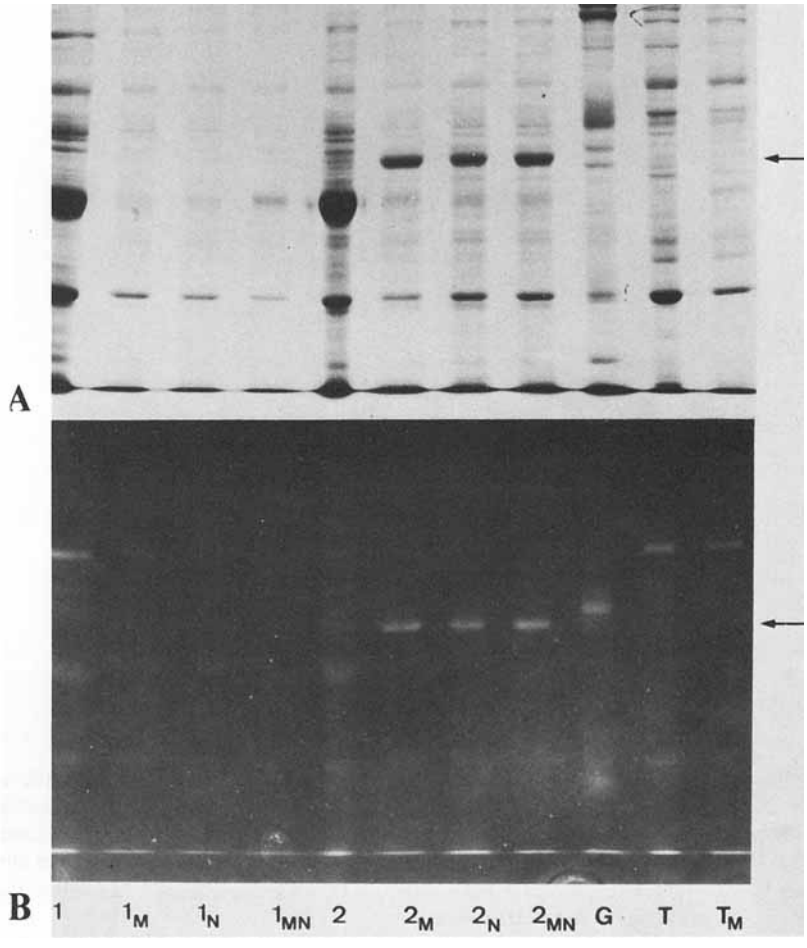

Fig.7. A) Protein composition of light and heavy particle fractions isolated from fresh thrombocyte-rich plasma. Plasma was processed as outlined in Figure 5 except that 20 mg/liter PMSF was added to the plasma prior to filtration. The light and heavy fractions following the dextran step gradient were divided in three equal parts, and anti-sera were added as indicated by subscript M or N (anti-M or anti-N). 1, supernate collected from all three agglutination assays from the light fraction; 2, supernate collected from all three agglutination assays from heavy fraction; 1_M, 1_N, 1_{MN}, agglutinated and pelleted material from the light fraction; 2_M, 2_N, 2_{MN}, agglutinated and pelleted material from the heavy fraction; G, 15 μgm protein of fresh human erythrocyte membranes; T, 15 μgm protein of thrombocyte membranes; T_M, thrombocyte membranes incubated with anti-M and pelleted.

B) Glycoprotein composition of light and heavy particle fraction isolated from the fresh thrombocyte-rich plasma.